Germination of spores by pressure

osides tested inosine strongly potentiated the other ribosides, n; these three were gher concentrations

es at 1 atm., did not

more or less directly mination caused by

rmination of

g incubation† in

Presence of OCDS‡

atm.	200 atm.
0	0
35.5	100
0	100
0	4
0	0
0	81.2
0	5.2
0	8
0	6
0	74
sured b	y recording

H 8.0) plus the indicated

e potentiation of pressure

lues (Fig. 2), pressure ntimetabolites known s showed that, just as *accelerating* germina*ng* germination as the

betyl alcohol, mercuric by pressure, suggested rather than by simple Rode & Foster, 1960).

Table 6. Potentiation of pressure germination of spores by ribosides, products of amino acid metabolism, and related compounds

Germination (%)* following incubation*

Addition	Concentration	Bacillus cereus		Bacillus coagulans		
		I atm.	200 atm.	I atm.	200 atm.	400 atm.
Control (no addition)		o	0	0	0	27
L-Alanine	I mM	25	100	40	92.5	94
	100 µm	I	100	11.2	81	92.5
Inosine	I mM	100	100	0	3	27
	100 µM	81	100	0	0	27
	30 µM	I	99	n.t.	n.t.	n.t. ‡
	ΙΟ μΜ	0	16.5	0	0	26
L-Alanine	100 µm)	87	100	13	82	95
plus inosine	ΙΟ μΜ					
Deoxyinosine	30 µm§	0	4	0	n.t.	24
Adenosine	30 µm§	0	20	0	n.t.	26
Deoxyadenosine	30 µm§	0	0	0	n.t.	26.5
Guanosine	30 µm§	0	9	0	n.t.	26
Deoxyguanosine	30 µm§	0	0	0	n.t.	25.5
Xanthosine	30 µм§	0	0	0	n.t.	26
Riboside breakdown products ¶	30 μm§	0	0	0	n.t.	24–29
Amino acid breakdown products ¶	30 μm§	0	0	0	0	n.t.

* Spores were activated before use (70°, 30 min.); germination was measured by recording the percentage of phase-dark spores.

 \dagger Spores were incubated for 30 min. at 30° (*Bacillus cereus*) or 40° (*B. coagulans*) in 0·I M-sodium phosphate (pH 8·0) plus the indicated additions.

‡ Not tested.

§ Concentrations were I mM for B. coagulans spores.

The following riboside and amino acid breakdown products and related metabolites were inactive: hypoxanthine, D-ribose, D-ribose 5-phosphate, 3-phosphoglycerate, 2-phosphoglycerate, pyruvate, ammonium (sulphate), nicotinamide adenine dinucleotide and nicotinamide adenine dinucleotide phosphate (oxidized and reduced forms), formate, acetate, acetyl phosphate, acetyl coenzyme A, coenzyme A, citrate, succinate, phosphopyruvate, adenosine diphosphate and adenosine triphosphate.

Unlike pressure on gaseous systems, hydrostatic pressure of the magnitude used here causes little compression of the liquid (water is compressed less than 4% at 1000 atm.) and therefore results in negligible changes in concentrations of solutes. Furthermore, the effectiveness of relatively low pressures (i.e. below 1000 atm.) in causing germination of certain spores suggested that pressure was not acting by causing denaturation of proteins or other macromolecules: such changes characteristically become pronounced at pressures well in excess of 1000 atm., and particularly at elevated temperatures. In fact a high proportion of spores survived pressures as high as 8000 atm. at room temperature (Sale *et al.* 1970). Laidler (1951) showed that at high temperatures the dominant effect of pressure on biological materials was on equilibria between natural and denatured forms of enzymes, whereas at lower temperatures the principal effect was on the rates of enzyme-substrate reactions. Pressures of the order of hundreds of atmospheres, at relatively low temperatures, usually affect the rates of reactions in which reactants and products have different molecular volumes.

Such an effect on a germination reaction which normally has a negligible rate at